Effect of Phytogenic Feed Mixture on Blood Hematology, Metabolites, Enzymes and Lipids in Broilers

Pooja Singh¹, Neeraj¹, Ramesh Pandey¹ and Amit Kumar Singh^{2*}

¹Department of Animal Husbandry & Dairying, Sam Higginbottom University of Agriculture, Technology and Sciences, Prayagraj, Uttar Pradesh, INDIA

²ICAR- Krishi Vigyan Kendra, Amihit, Jaunpur, Acharya Narendra Dev University of Agriculture & Technology, Ayodhya, Uttar Pradesh, INDIA

*Corresponding author: AK Singh; E-mail: amitkumarsingh5496@gmail.com

Received: 01 Aug., 2022

Revised: 04 Sept., 2022

Accepted: 10 Sept., 2022

ABSTRACT

Phytogenic feed additives have been used traditionally in medicine preparations and in the preparation of culinary food dishes. However, in recent years it has attracted researchers for their use as feed additives in broilers. This study was organized to access the effect of dietary supplementation of phytogenic mixture containing powder of *Allium sativum* bulbs, *Ocimum sanctum* leaves and *Nigella sativa* seeds in 1:1:1 ratio. For this study, three comparable groups were formed viz. control (non supplementation), T_1 (supplementation @0.25%) and T_2 (supplementation @0.50%). A total of 48 broiler chicks were randomly distributed in three groups, each group containing 16 chicks of same strain and body weight. The experiment lasted for a period of 28 days. Statistical analysis of data revealed a significant (*P*<0.001) change in blood lipid profile of three groups. T_1 and T_2 groups had significantly lower (*P*<0.001) levels of triglycerides, total cholesterol, HDL cholesterol, LDL cholesterol and Non-HDL cholesterols (*P* = 0.05). However, a non-significant (*P*>0.05) difference among three groups on blood hematology (Hemoglobin, WBC, RBC, PCV and Platelet count) was observed. Study showed that there was statistically similar (*P*>0.05) concentrations of blood metabolites (Glucose, Total protein, Albumin, Globulin) and statistically similar (*P*>0.05) levels of blood enzyme (Bilirubin, serum AST, serum ALT and serum ALP) among three groups. Based on the findings of this study, conclusively, it can be said that dietary supplementation of phytogenic mixture containing powder of *Allium sativum* bulbs, *Ocimum sanctum* leaves and *Nigella sativa* seeds in 1:1:1 ratio may favorably lower the levels of blood lipid without adversely affecting the normal physiology of broilers.

HIGHLIGHTS

- Phytogenic mixture improves physiological working of broilers.
- Phytogenic mixture feeding lowered blood cholesterol in broilers.
- Phytogenic mixture didn't affect hematology, metabolites and enzymes levels.

Keywords: Broilers, blood lipid, blood hematology, blood metabolites, blood enzymes, phytogenic mixture

As people are getting aware about the importance of quality protein which may biologically become available to their body easily. Many people are shifting to non-vegetarian foods. Consumption of meat has increased under Indian conditions and according to the survey of comptroller general of India, more than two third of youths of India consume meat (Singh *et al.*, 2021a; Singh *et al.*, 2021). Poultry meat has been seen as fastest growing

and consumed meat around the world as it is cheaper, provides quality meat, among several other attributes (Singh *et al.*, 2021b). However, consumption of meat in

How to cite this article: Singh, P., Neeraj, Pandey, R. and Singh, A.K. (2022). Effect of Phytogenic Feed Mixture on Blood Hematology, Metabolites, Enzymes and Lipids in Broilers. *J. Anim. Res.*, **12**(05): 713-719.

Source of Support: None; Conflict of Interest: None



India is around 5 kg per person whereas; consumption of meat in United States of America and Australia is around 100kg per person annually (Ritchie and Roser, 2017; Nagar et al., 2020; Nagar et al., 2021). Cultivable land for agriculture is limited for production of food grains and other nutrient providing sources (Arora, 2019; Vianna et al., 2022). Production of poultry requires a high amount of concentrates feeding which directly competes for the resources available for production of food items for human (Mengesha, 2012; Thakur et al., 2020a; Thakur et al., 2020b). Hence, researchers are continuously engaged in finding economical feed additives for broilers which may enhance broiler production (Amsathkumar et al., 2019). With the ban of antibiotic feed additives in broiler feed has pushed researchers for alternative feed additives which promote broiler production (Ayalew et al., 2022). Among such alternative feed additive the emerging and economical feed additives is phytogenic mixture as is it environment friendly, non-toxic, doesn't have residual effects (Amsathkumar et al., 2019).

Phytogenic feed additives may be defined as ingredients which are derived from plant parts such as leaves, bulbs, roots, etc. incorporated into the animal feed to enhance animal's performances by the virtue of their ability to improve digestibility, nutrient assimilation and destroying pathogens found in animal digestive tracts and additionally they improve the quality products from the animals fed with phytogenic feed mixture (Kamel, 2001; Balunas and Kinghorn, 2005; Windisch et al, 2008). Among several phytogenic feed additives, Allium sativum, Ocimum sanctum and Nigella sativa have gained much attention from scientific community for their growth and health promoting attributes in broilers as they as economical and easily available throughout the world (Bihari et al., 2010; Abu-Al-Basal, 2011; Khan et al., 2012; Palani et al., 2014; Puvaca et al., 2015; Prajapat et al., 2018). Several studies suggest that combination of two or more phytogenic ingredients synergistically improves the performance of poultry birds (Amsathkumar et al., 2019, Ali et al., 2014).

Much work has been done to examine the effects of dietary supplementation of *Allium sativum*, *Ocimum sanctum* and *Nigella sativa* individually on blood profile of broilers. However, to the knowledge of authors, no experiment has been structured to observe the combined effect of *Allium sativum*, *Ocimum sanctum* and *Nigella sativa* mixture on blood profile of broilers. Therefore, the present study was structured with an objective to investigate the effect of phytogenic mixture containing *Allium sativum*, *Ocimum sanctum* and *Nigella sativa* on blood lipid profile, hematology, metabolites and enzyme concentrations of broilers.

MATERIALS AND METHODS

Site of the study

The present experiment was carried out in small animal laboratory of the Department of Animal Husbandry & Dairying, SHUATS, Prayagraj-211007.

Pre-experimental preparations

Prior to placement of the chicks in the cages, the entire poultry house and its premises were thoroughly cleaned with water and disinfected followed by fumigation with formalin and potassium permanganate (KMnO₄ @0.02%). To ensure the maintenance of the bio-safety security of the house, a disinfectant (lime powder) was always provided as a dip at the entrance of the house.

Preparation of phytogenic mixture

Garlic (cloves), Tulsi (leaves) and black cumin (seeds) were purchased from the local market. All three ingredients were procured fresh from the local market. The bulb portion of garlic was peeled followed by slicing into small pieces, fresh Tulsi leaves were washed with clean water and seeds of black cumin seeds were taken together and dried at 40°C for 24 hours in hot air oven. Dried samples were converted to powder using electric blender and then transferred to separate and labeled air tight vessels and kept in a cool dry place away from sunlight till further usage.

Experimental animals and management

A total of 48 straight run Cobb 400 broiler chicks were purchased from a corporate hatchery firm. Chicks were then administered anti-stress (sugared solution and multivitamins) after their arrival and were distributed according to the treatment groups and identified individually by means of wing tags. Battery type cages were used for rearing birds and they were provided with standard farm managemental practices providing 1 sq. ft/bird space from day-old to four weeks of age. Fresh feed and clean water were served *ad libitum*. Birds were reared under the same environmental conditions. During the brooding, chicks were kept under the maintained temperature at 35°C during the first week, followed by lowering the temperature by 3°C every week till temperature of room was adjusted to 25°C in following days. One bulb of 100 watt was provided in each cage for light and to maintain the temperature in the room. The experimental period was for 28 days from starting week of February 2022 to second week of March 2022.

Experimental design and treatments

The birds were assigned into three comparable treatment groups. Each treatment group had 16 birds comprising 4 replicates in each group with 4 birds per replicate in a complete randomized design. Each group was fed with corresponding experimental diet—

- **T**₀: Chicks were provided with standard ration as per NRC, 2007 standards, without any supplement.
- $\Box \quad T_1: \text{ Standard ration supplemented with } 0.25\% \text{ of } phytogenic mixture per kg of feed.}$
- $\Box \quad T_2: \text{ Standard ration supplemented with } 0.50\% \text{ of } phytogenic mixture per kg of feed.}$

Phytogenic mixture contained Garlic, Tulsi and Black Cumin powder mixed in equal proportions 1:1:1 and then supplemented in the desired amount in respective treatment groups. The broiler starter diet contained 22% crude protein (CP) with 3, 000 ME kcal/kg whereas, the broiler finisher diet had 20% CP and 3, 150 ME kcal/ kg in accordance with NRC standards. The feeding and watering was made available *ad-libitum* to the birds.

Blood hematology and biochemical components

Towards the completion of this study, blood samples were collected aseptically from the wing vein of each bird using disposable syringe into sterilized tubes. Serum samples were obtained from blood samples as per Sitohy *et al.* (2013), Ashour *et al.* (2020). Blood heamatology, including red blood cells (RBC) count, total white blood cells (WBC) count, hemoglobin (Hb) concentration,

packed cell volume (PCV), platelet count by standard procedures. Serum samples were analyzed using spectrophotometer for Glucose (G), total protein (TP), albumin (Alb), Globulin (Glb), serum enzymatic activities of alanine amino transferase (ALT) and aspartate amino transferase (AST), total bilirubin, alkaline phosphatase (ALP), total cholesterol (TC), triglycerides (TG), lowdensity lipoprotein (LDL) and high-density lipoprotein (HDL) were estimated through commercially available standard diagnostic kits as followed by Sinha *et al.* (2017).

 Table 1: Ingredient and nutrient composition of experimental diet (%DM)

Ingredients (%)	Broiler starter (0 – 21 day)	Broiler finisher (22 – 42 days)				
Corn	53.53	<u>(22 – 42 days)</u> 59.58				
Soyabean meal (44 %CP)	38.95	33.33				
Monodibasic Phosphate	1.44	1.22				
Limestone	1.34	1.37				
Vegetable oil	3.85	3.53				
Salt	0.40	0.41				
DL- Methionine	0.208	0.215				
L-Lysine- HCL	0.128	0.196				
Choline HCL (60%)	0.06	0.05				
Mineral- Vitamins premix	0.01	0.01				
Total	100	100				
Calculated Nutrients						
Crude protein %	22	20				
ME, Kcal/kg	3000	3,150				
Calcium %	0.91	0.88				
Available phosphorus %	0.40	0.36				
Sodium %	0.21	0.22				
Chloride %	0.26	0.28				
Digestible Lys. %	1.17	1.08				
Digestible Met. %	0.48	0.49				
Digestible Met + Cys %	0.82	0.78				
Digestible Thr, %	0.79	0.73				
Choline, mg/kg	1,422	1,312				

Statistics and data analysis

All data collected were analysed meticulously for analaysis of variance (ANOVA) using Statistical Analysis System software of IBM (SPSS 22). Microsoft Excel was used for statistics and graphical representations. Duncan



Multiple Range Test was performed for determining the significant differences among different groups. Difference was considered significant when the value of P was lower or equal to 0.05.

RESULTS AND DISCUSSION

From the perusal of statistically analyzed data (Table 2), non-significant difference (P>0.05) was observed in hematological parameters (Hemoglobin, RBC, WBC, PCV and platelet count), blood metabolites (Glucose, total protein, albumin and globulin) and blood enzymes (Bilirubin, SGOT, SGPT and Alkaline Phosphatase). However, significant (P<0.001) difference was reported for blood cholesterol, triglycerides, HDL Cholesterol and LDL in treatment groups T₁ and T₂ birds than control group in this experiment. Nevertheless, T₁ and T₂ groups differed non-significantly (P<0.05) for blood cholesterol and LDL

cholesterol while Triglycerides and HDL cholesterol values differ significantly (P<0.05) in group T₁ and T₂.

Significantly non-different (P<0.05) blood metabolites, blood enzymes and hematological parameters in treatment group birds show that the supplementation of phytogenic feed had no adverse effect on the normal physiology of birds. The significant difference (P<0.001) recorded in the blood cholesterol, triglycerides, HDL cholesterol and LDL in treatment groups T₁ and T₂ birds as compared to control group. Similar result was reported by Bamidele and Adejumo (2012) who found that the blood cholesterol and HDL cholesterol were significantly reduced in the treatment group. Lowest cholesterol in the treatment group was reported by Borgohain *et al.* (2019) by supplementing garlic powder @ 1 % and 1.5% of in the diet of broilers. Ismail *et al.* (2021) also analyzed and reported that total cholesterol and LDL cholesterol were significantly

Table 2: Least square means of indices of blood hematology, metabolites, enzymes and lipids in broilers

Parameters	T	T ₁	T ₂	SEM	Statistical Significance			
Blood Hematology								
Haemoglobin (g/dl)	9.89	9.82	9.76	0.09	NS <i>P</i> = 0.862			
WBC (10 ³ /ul)	75.05	74.88	74.82	0.17	NS $P = 0.840$			
RBC (10 ⁶ /ul)	2.28	2.27	2.24	0.02	NS $P = 0.661$			
PCV (%)	30.28	30.04	30.51	0.27	NS $P = 0.778$			
Platelet Count (10 ³ / cu mm)	10.94	10.88	10.87	0.11	NS $P = 0.968$			
	Blo	od Metabolit	es					
Glucose (mg/dl)	224.12	223.75	224.38	1.19	NS <i>P</i> = 0.977			
Total Protein (g/dl)	2.78	2.81	2.77	0.02	NS $P = 0.700$			
Albumin (gm/dl)	1.35	1.36	1.36	0.02	NS <i>P</i> = 0.990			
Globulin (gm/dl)	1.38	1.38	1.39	0.01	NS $P = 0.914$			
	В	lood Enzyme	5					
Bilirubin-Total (mg/dl)	0.29	0.28	0.29	0.01	NS <i>P</i> = 0.902			
Serum AST/SGOT (U/L)	235.94	235.50	240.25	1.47	NS $P = 0.354$			
Serum ALT/SGPT (U/L)	8.50	8.48	8.56	0.07	NS $P = 0.883$			
Alkaline Phosphatase (U/L)	2025.44	2026.12	2025.25	9.00	NS <i>P</i> = 0.999			
		Blood lipids						
Cholesterol (mg/dl)	132.31ª	118.19 ^b	117.69 ^b	0.67	S <i>P</i> = 0.000			
Triglycerides (mg/dl)	61.69 ^a	54.19 ^b	52.06 °	0.37	S <i>P</i> = 0.000			
HDL Cholesterol (mg/dl)	90.38 ^a	71.12 ^b	74.81 ^c	0.65	S P = 0.000			
Non-HDL Cholesterol (mg/dl)	41.94	47.06	42.88	0.89	S <i>P</i> = 0.05			
LDL Cholesterol (mg/dl)	28.88 ^a	22.50 ^b	23.50 ^b	0.34	S P = 0.000			

Least Square Means bearing different superscripts differ significantly (P<0.05) column wise in same row.

reduced by adding garlic powder in the diet of broilers. The present results were in agreement with Miraghaee *et al.* (2011) who reported that total serum cholesterol and triglycerides were decreased in group fed 1% black cumin seed than control. (Shewita and Taha, 2011; Nasir, 2005; Al-Beitawi and El-Ghousein, 2008; Fadlalla *et al.*, 2008) also reported the decrease in serum total cholesterol and serum triglyceride levels with supplementation of black cumin seed in broilers ration.

In earlier study, Fadlalla et al. (2008) also reported the similar findings for blood serum protein, albumin and globulin level which did not get affected significantly by adding 0.3% graded garlic in the treatment group. No significance difference in the blood hematological parameters was found by (Fadlalla et al., 2008; Khatun et al., 2013) in the treatment group as compare to control by supplementing different levels of tulsi and neem extract in drinking water of the birds. This result was similar to our findings in which the blood hematological parameters did not get significantly affected by treatments. However, there are some studies which reported contrary results for hematological parameters (PCV, Hb, RBC, and WBC) which were found to be significantly (P < 0.05) affected by lemon grass - garlic extracts (CLGE) augmentation in the broiler's diet (Alam et al., 2014). However, (Fadlalla et al., 2008; Alagbe and Oluwafemi, 2019; Jahejo et al., 2019) showed that WBC was significantly increased in the treatment groups by supplementation of phytogenic mixture. The reason behind this phenomenon may be concentrations of cholesterol gets altered with balance between dietary inputs and body synthesis. Hence, if any feed material which may change the working of cells of body, it might play a role in changes in the levels of cholesterol. The reduced levels of plasma cholesterol may be due to supplement of one of the ingredients, Nigella sativa which may stimulate the cholesterol excretion into the intestine (Khodary et al., 1996). El-Alfy et al. (1975) investigated Nigella sativa oil has cholorelic effect that may elevate bile flow. Nevetheless, bile flow is an emulsifying agent which activates pancreatic lipase and finally contributes in digestion and absorption of fat and fat- soluble vitamins (Crossland, 1980).

CONCLUSION

Based on the findings of this study, it may be concluded that dietary supplementation of phytogenic mixture containing

powder of *Allium sativum* bulbs, *Ocimum sanctum* leaves and *Nigella sativa* seeds in 1:1:1 ratio may favorably lower the levels of blood lipids without adversely affecting the normal physiology of broilers. Though, the number of chicks was limited in our study, hence, it is recommended to conduct further studies on large number of birds.

ACKNOWLEDGEMENTS

Authors have deep regards towards Vice Chancellor, Sam Higginbottom University of Agriculture, Technology and Sciences, Prayagraj, India for academic support and provision of necessary facilities for conducting this study. Authors are also thankful towards Head of Department of Animal Husbandry & Dairying, SHUATS, India for their guidance in academic prospects of corresponding author.

REFERENCES

- Abu-Al-Basal, M.A. 2011. Influence of *Nigella sativa* fixed oil on some blood parameters and histopathology of skin in staphylococcal-infected BALB/c mice. *Pakistan J. Biol. Sci.*, 14: 1038-1046.
- Alagbe, J.O. and Oluwafemi, R.A. 2019. Performance and haematological parameters of broiler chicks gives different levels of dried lemon grass (*Cymbopogon citratus*) and garlic (*Allium sativum*) extract. *Res. Agric. Vet. Sci.*, **3**: 102-111.
- Alam, M.N., Uddin, J., Akter, M.T.D., Moni, M., Alom, I.Z., Rahman, F.A. and Al Noman, M.A. 2014. Broiler production by using polyherbal medication (neem, nishyinda, tulsi and turmeric extract). *Int. J. Innov. Appl. Stud.*, **9**: 1161-1175.
- Al-Beitawi, N. and El-Ghousein, S.S. 2008. Effect of feeding different levels of *Nigella sativa* seeds (black cumin) on performance, blood constituents and carcass characteristics of broiler chicks. *Int. J. Poultry Sci.*, 7: 715-721.
- Ali, S., Mukhtar, M., Manzoor, S., Hssain, Z., Ali, A., Tabassum, R., Imran, M., Amer, M.Y. and Bhatti, N. 2014. Effect of garlic, black seed and turmeric on the growth of broiler chicken. *Pakistan J. Nut.*, **13**: 204.
- Amsathkumar, L., Jadhav, S.E., Pattanaik, A.K. and Dutta, N. 2019. Nutrient utilization and performance of endotoxin exposed kids supplemented with phytogenic feed additive. *Anim. Nut. Feed Technol.*, **19**: 371-383.
- Arora, N.K. 2019. Impact of climate change on agriculture production and its sustainable solutions. *Envir. Sust.*, 2: 95-96.
- Ashour, E.A., Abd El-Hack, M.E., Swelum, A.A., Osman, A.O., Taha, A.E., Alhimaidi, A.R. and Ismail, I.E. 2020. Does the dietary graded levels of herbal mixture powder impact



growth, carcass traits, blood indices and meat quality of the broilers?, *Italian J. Anim. Sci.*, **19**: 1228-1237.

- Ayalew, H., Zhang, H., Wang, J., Wu, S., Qiu, K., Qi, G., Tekeste, A., Wassie, T. and Chanie, D. 2022. Potential feed additives as antibiotic alternatives in broiler production. *Front. Vet. Sci.*, **9**: 916473.
- Balunas, M.J. and Kinghorn, A.D. 2005. Drug discovery from medicinal plants. *Life Sci.*, 78: 431-441.
- Bihari, C.G., Shankar, N.B., Kumar, J.P., Keshari, P.S. and Ellaiah, P. 2010. Phytochemical investigation and screening for anthelmintic activity of leafy extracts of various *Ocimum* (Tulsi) species. *J. Pharmacy Res.*, **3**: 2140-2141.
- Borgohain, B., Mahanta, J.D., Sapcota, D., Handique, B. and Islam, R. 2019. Effect of feeding garlic (*Allium sativum*) on haematological, serum biochemical profile and carcass characteristics in broiler chicken. *Int. J. Curr. Microbiol. Appl. Sci.*, 8: 492-500.
- Crossland, J. 1980. *Lewiss Pharmacology*. 5th Ed, Churchill Livingston, London, N.Y., pp. 656-657.
- El-Alfy, T.S., El-Fatatry, H.M. and Toama, M.A. 1975. Isolation and structure assignment of an antimicrobial principle from the volatile oil of *Nigella sativa L. Pharmazie*, **30**: 109.
- Fadlalla, I.M.T., Mohammed, B.H. and Bakhiet, A.O. 2010. Effect of feeding garlic on the performance and immunity of broilers. *Asian J. Poultry Sci.*, 4: 182-189.
- Ismail, I.E., Alagawany, M., Taha, A.E., Puvača, N., Laudadio, V. and Tufarelli, V. 2021. Effect of dietary supplementation of garlic powder and phenyl acetic acid on productive performance, blood haematology, immunity and antioxidant status of broiler chickens. *Anim. Biosci.*, 34: 363.
- Jahejo, A.R., Rajput, N., Wen-Xia, T., Naeem, M., Kalhoro, D.H., Kaka, A. and Jia, F.J. 2019. Immunomodulatory and growth promoting effects of basil (*Ocimum basilicum*) and ascorbic acid in heat stressed broiler chickens. *Pakistan J. Zool.*, **51**: 801.
- Kamel, C. 2001. Natural plant extracts: classical remedies bring modern animal production solutions. *Cahiers Options Méditerranéennes*, 54: 31-38.
- Khan, S.H., Ansari, J., Haq, A.U. and Abbas G. 2012. Black cumin seeds as phytogenic product in broiler diets and its effects on performance, blood constituents, immunity and caecal microbial population. *Italian J. Anim. Sci.*, **11**: 438-444.
- Khatun, S., Mostofa, M., Alom, F., Uddin, J., Alam, M.N. and Moitry, N.F. 2013. Efficacy of tulsi leaves and neem leaves extract in broiler production. *Bangladesh J. Vet. Med.*, 11: 1-5.
- Khodary, R.M., EL-Ezzawy, M.H. and Hamdy, I.R. 1996. Effect of *Niglla sativa* on egg production, hatchability percentage

and some biochemical values in laying hen with references to fertility in cockerels. Proc of 7th Sci. Cong., Fac. Vet. Med., Assuit Univ., 17-19 Nov. Ass. Egypt, pp. 91-106.

- Mengesha, M. 2012. The issue of feed-food competition and chicken production for the demands of foods of animal origin. *Asian J. Poultry Sci.*, 6: 31-43.
- Miraghaee, S.S., Heidary, B., Almasi, H., Shabani, A., Elahi, M. and Nia, M.H.M. 2011. The effects of *Nigella sativa* powder (black seed) and *Echinacea purpurea* (*l.*) Moench extract on performance, some blood biochemical and hematological parameters in broiler chickens. *African J. Biotechnol.*, 10: 19249-19254.
- Nagar, A., Neeraj, Pandey, R. and Singh, A.K. 2020. Influence of dietary supplementation of Shatavari (*Asparagus racemosus*) and Ashwagandha (*Withania somnifera*) root powder on feed intake and body weight performance in caged broilers. *J. Entomol. Zool. Stud.*, 8: 592-597.
- Nagar, A., Neeraj, Pandey, R., Singh, A.K. and Thakur, R. 2021. Impact of dietary supplementation of Shatavari (*Asparagus racemosus*) and Ashwagandha (*Withania somnifera*) root powder on performances in broilers. J. Anim. Res., 11(2): 333-339.
- Nasir, Z. 2005. Effect of kalongi (*Nigella sativa*) seeds on egg production and quality in white leghorn layers. J. Anim. Plant Sci., 15: 22-24.
- Palani, S., Joseph, N.M., Tegene, Y. and Zacharia, A. 2014. Garlic - a concise review. *Int. J. Pharmacog.*, 1: 691-95.
- Prajapat, U.K., Jain, D., Dhuria, R.K., Sharma, T., Bothra, T., Nehra, R. and Kumar, M. 2018. Effect of dietary supplementation of tulsi (*Ocimum sanctum*) leaf powder and fenugreek (*Trigonella foenum graecum l.*) seed powder on growth performance in broilers. *Vet. Pract.*, 19: 1.
- Puvača, N., Ljubojević, D., Kostadinović, L.J., Lukač, D., Lević, J., Popović, S. and Đuragić, O. 2015. Spices and herbs in broilers nutrition: Effects of garlic (*Allium sativum* L.) on broiler chicken production, *World's Poultry Sci. J.*, 71: 533-538.
- Ritchie, H. and Roser, M. 2017. "Meat and Dairy Production". Published online at OurWorldInData.org. Retrieved from: https://ourworldindata.org/meat-production
- Shewita, R.S. and Taha, A.E. 2011. Effect of dietary supplementation of different levels of black seed (*Nigella* sativa 1.) on growth performance, immunological, hematological and carcass parameters of broiler chicks. World Acad. Sci. Eng. Technol., **77**: 788-794.
- Singh, A.K., Debbarma, A., Baishya, A., Sarkar, D., Mohanta, K.P. and Anil. 2021b. Insights of Improved Backyard Poultry Farming in India with Special Reference to Hilly Regions: A Review. *Int. J. Livest. Res.*, **11**: 1-16.

Journal of Animal Research: v. 12, n. 05, October 2022

- Singh, A.K., Singh, P., Yadav, S.K. and Anil. 2021a. Role of backyard poultry in sustainable rural livelihood under Indian perspectives. *Scientific India*, 9: 41-43.
- Singh, P., Singh, A.K., Neeraj and Pandey, R. 2021. Use of insects as a protein source for broiler production under Indian conditions. *Scientific India*, 9: 10-12.
- Sinha, S., Muzamil, S., Ahmad, B., Rehman, M.U. and Quadri, A. 2017. Ameliorative effect of aloe vera supplementation in poultry feed. J. Anim. Res., 7: 85-90.
- Sitohy, M., Osman, A., Gharib, A., Chobert, J.M. and Haertl, T. 2013. Preliminary assessment of potential toxicity of methylated soybean protein and methylated b-lactoglobulin in male Wistar rats. *Food. Chem. Toxicol.*, **59**: 618–625.
- Thakur, R., Neeraj, Pandey, R., Singh, A.K. and Nagar, A. 2020a. Effect of cardamom and ginger powder supplementation on body weight gain and feed efficiency in caged broilers. *Int. J. Curr. Microbiol. App. Sci.*, 9: 2159-2168.

- Thakur, R., Neeraj, Pandey, R., Singh, A.K. and Nagar, A. 2020b. Effect of cardamom and ginger powder supplementation on growth performance in caged broilers. *Int. J. Livest. Res.*, 10: 155-162.
- Viana, C.M., Freire, D., Abrantes, P., Rocha, J. and Pereira, P. 2022. Agricultural land systems importance for supporting food security and sustainable development goals: A systematic review. *Sci. Total Environ.*, **806**: 150718.
- Windisch, W., Schedle, K., Plitzner, C. and Kroismayr, A. 2008. Use of phytogenic products as feed additives for swine and poultry. J. Anim. Sci., 86: E140-E148.