

PLANT PATHOLOGY

Efficacy of different Fungicide against *Alternaria solani* Caused Early Blight Disease of Tomato under *in vitro* Condition

Sunita Dhaka^{1*} and Anand Choudhary²

¹Department of Plant Pathology, Jawaharlal Nehru Krishi Vishwa Vidyalaya, Jabalpur, M.P., India ²Department of Plant Pathology, Swami Keshwanand Rajasthan Agriculture University, Bikaner, Rajasthan, India

*Corresponding author: imsunita1997@gmail.com (ORCID ID: 0000-0003-4808-1746)

 Paper No. 958
 Received: 14-12-2021
 Revised: 27-02-2022

Accepted: 09-03-2022

ABSTRACT

Tomato (*Solanum lycopersicum* L.) is one of the most popular vegetable crops. It is affected by several diseases among them early blight caused by *Alternaria solani* is most destructive foliar diseases of tomato, the present study was conducted to evaluate *in vitro* efficacy of different fungicides with four different concentrations (250, 500, 750 &1000 ppm). Among them highest percent inhibition was found in Propiconazole (90.19%) at 1000 ppm concentration after 120 hrs followed by Tebuconazole (86.52%), Mancozeb (80.63%) and Captan + Hexaconazole (79.40%) against *A. solani*. and Moderately growth was found in Hexaconazole (78.17%), Thiram (76.64%) Captan (74.73%) and Carbendazim (65.54%). Lowest percent inhibition was observed in Copper oxychloride (42.65%) against control (65.30mm).

HIGHLIGHTS

- Early blight disease is painful nerve to tomato growing farmer.
- Propiconazole and Tebuconazole were the best fungicides for effectively managing early blight disease of tomato.

Keywords: Fungicides, tomato, early blight, Alternaria solani, mycelial inhibition

Tomato (*Solanum lycopersicum* L.) is one of the most popular vegetable crops cultivated extensively all over the world for its edible fruits. It belongs to family *Solanaceae* and originated from the Andes, in what is now called Bolivia, Central America (Peru), Chili and Ecuador where they grow wild (Mehta 2017). It is a traditional vegetable crop commercially cultivated in India and ranks second in the production of tomato after China. Tomato is a rich source of several nutrients, vitamins (A, B, C) and minerals and antioxidants such as carotenoids. Well drained sandy loam soil with high level of contents is suitable for tomato cultivation.

The crop occupies 8.85 thousand/ha Land in India with 19696 m. tones production and 24.4 m. tones/ ha productivity during 2016-17 (Anon 2017).

Tomato can be affected by many diseases during growing season and is vulnerable for biotic and

abiotic stresses. The biotic contributors such as fungi, bacteria, virus and nematodes causing major losses to the crop (Bost 2013; Gleason and Edmunds, 2006). Two common and devastating disease of tomato are early blight & *Fusarium* wilt caused by *Alternaria solani* and *Fusarium oxysporum f. sp. lycopersici* respectively and which produces leaf spots, stem canker and fruit rots, but the foliar phase is the most common & destructive part of the disease. (Maiero and Barksdale 1989). Early blight, incited by *Alternaria solani* (Ellis & Martin) Jones and Grout is one of the most destructive foliar diseases of tomato, causing significant reduction

How to cite this article: Dhaka, S. and Choudhary, A. (2022). Efficacy of different Fungicide against *Alternaria solani* Caused Early Blight Disease of Tomato under *in vitro* Condition. *Int. J. Ag. Env. Biotech.*, **15**(01): 61-65.

Source of Support: None; Conflict of Interest: None



in the quality and quantity of fruit yields (Tewari and Vishunavat 2012). The disease is responsible to cause 35.14 percent yield losses under open field and 33.94 to 69.28 Percent in poly house (Soni et al. 2017). This disease causes losses in fruit yield from 50-80% (Mathur and Shekhawat 1986). The disease is characterized by the appearance of brown to dark brown necrotic spots with concentric rings on foliage, stem and fruits (Chouhan et al. 2015) and oval or angular in shape ranging from 0.3 to 0.4 cm diameter with usually narrow chlorotic zone around the spot (Walker 1952). The first symptom of the Early Blight appears on the lower senescent leaves as dark necrotic lesions. The disease process upward as the plant become older. The infection on fruit causes dark, sunken, leathery and purple lesions on the stem-end. These lesions expand to a significant size and extend deep into the flesh of the fruit. Infected fruits mostly drop prematurely, and those reaching to the maturity also become unmarketable (Chaerani and Voorrips 2006). A. solani infect all the above ground parts of the plant at all stage of growth and development (Peralta et al. 2005; Verma et al. 2007). The plants are more susceptible to infection by the pathogen during fruiting period (Cerkauskas 2005; Momel and Pemezny 2006). Keeping in view the importance of the disease, the present study was conducted to evaluate in vitro efficacy of different fungicides against Alternaria solani.

MATERIALS AND METHODS

In vitro evaluation of fungicides

Nine fungicides were evaluated under laboratory conditions against A. solani by Poisoned Food Techniques (Nene and Thapliyal 1973). Observation on mycelial growth was recorded after 120 hrs incubation.

| Table 1: Fungicides used against A. solani |
|--|
|--|

| Treatment | Fungicides | Concentration (ppm) |
|----------------|--------------------|---------------------|
| T ₁ | Mancozeb | 250,500,750,1000 |
| T ₂ | Copper oxychloride | 250,500,750,1000 |
| T ₃ | Carbendazim | 250,500,750,1000 |
| T_4 | Tebuconazole | 250,500,750,1000 |
| T ₅ | Propiconazole | 250,500,750,1000 |
| T ₆ | Captan | 250,500,750,1000 |
| T ₇ | Thiram | 250,500,750,1000 |

| T ₈ | Captan + Hexaconazole | 250,500,750,1000 |
|-----------------|--------------------------|------------------|
| T ₉ | Hexaconazole | 250,500,750,1000 |
| T ₁₀ | Control | _ |
| | | |

Poison Food Technique

Required quantity of each fungicide under study was mixed thoroughly in sterilized 100 ml PDA media filled in 250 ml flask separately under aseptic condition. The medium was supplemented with streptomycin sulphate @ 50 ppm to prevent bacterial contamination. The poisoned medium was then poured in sterilized petri plates (20 ml) and allowed it to solidify. Mycelium discs of 5 mm size from after 120 hrs incubation was cut by a sterile cork borer and one such disc was placed at the center of each agar plate. The plate without any fungicide served as control. Three replications were maintained for each concentration. Such plates were incubated at room temperature and the radial growth was measured when fungus attained maximum growth in control plates. Percent inhibition of mycelial growth over untreated control was calculated by applying the formula given by Vincent (1947).

RESULTS AND DISCUSSION

Nine different fungicides viz., Mancozeb, Copper oxychloride, Carbendazim, Tebuconazole, Propiconazole, Captan, Thiram, Captan + Hexaconazole and Hexaconazole were evaluated on mycelial growth of A. solani at different 250, 500, 750, 1000 ppm concentrations.

The data presented table 2, Fig. 1 and Plate 1 indicated that, highest percent inhibition was found in Propiconazole (90.19%) at 1000 ppm concentration after 120 hrs followed by Tebuconazole (86.52%), Mancozeb (80.63%) and Captan + Hexaconazole (79.40%) against A. solani. and Moderately growth was found in Hexaconazole (78.17%), Thiram (76.64%) Captan (74.73%) and Carbendazim (65.54%). Lowest percent inhibition was observed in Copper oxychloride (42.65%) against control (65.30 mm). Whereas Tebuconazole and Propiconazole inhibited 71.16% mycelial growth at 250 ppm followed by Captan + Hexaconazole (56.62%), Thiram (56.32%).

The observations are similar observations were also recorded by to Sahu et al. (2013), Chouhan

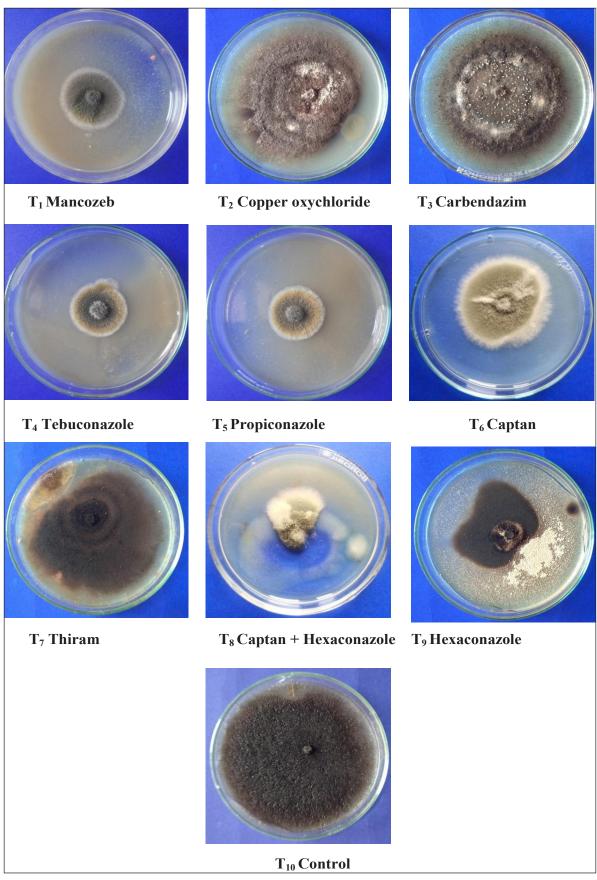


Plate 1: Efficacy of different fungicides against Alternaria solani

A



| Treatment | T | Radial Mycelial Growth (mm) Percent inhibition | | | | | | | |
|-----------------|-----------------------|---|-------|---------|-------|---------|-------|----------|-------|
| | Fungicides | 250 ppm | | 500 ppm | | 750 ppm | | 1000 ppm | |
| | | RMG | PI | RMG | PI | RMG | PI | RMG | PI |
| T ₁ | Mancozeb | 22.65 | 65.31 | 18.83 | 71.16 | 14.50 | 77.79 | 12.65 | 80.63 |
| T_2 | Copper oxychloride | 55.52 | 15.28 | 48.25 | 26.11 | 45.12 | 30.90 | 37.45 | 42.65 |
| T ₃ | Carbendazim | 46.52 | 28.75 | 33.62 | 48.51 | 27.45 | 57.96 | 22.5 | 65.54 |
| T ₄ | Tebuconazole | 18.83 | 71.16 | 15.97 | 75.54 | 12.12 | 81.44 | 8.52 | 86.52 |
| T ₅ | Propiconazole | 18.83 | 71.16 | 15.97 | 75.54 | 10.15 | 84.45 | 6.40 | 90.19 |
| T ₆ | Captan | 29.65 | 54.59 | 20.67 | 68.35 | 18.05 | 72.35 | 16.50 | 74.73 |
| T ₇ | Thiram | 28.52 | 56.32 | 18.52 | 71.63 | 16.50 | 74.73 | 15.25 | 76.64 |
| T ₈ | Captan + Hexaconazole | 28.33 | 56.62 | 19.43 | 70.25 | 15.45 | 76.34 | 13.45 | 79.40 |
| T ₉ | Hexaconazole | 29.52 | 54.79 | 20.52 | 68.57 | 17.85 | 72.66 | 14.25 | 78.17 |
| T ₁₀ | Control | 65.30 | | 65.30 | _ | 65.30 | _ | | |
| | SE(m)± | | | 0.51 | | 0.55 | | | |
| | CD (P=0.05) | | | 1.42 | | 1.42 | | | |

Table 2: Efficacy of different fungicides against Alternaria solani

*Mean of three replications.

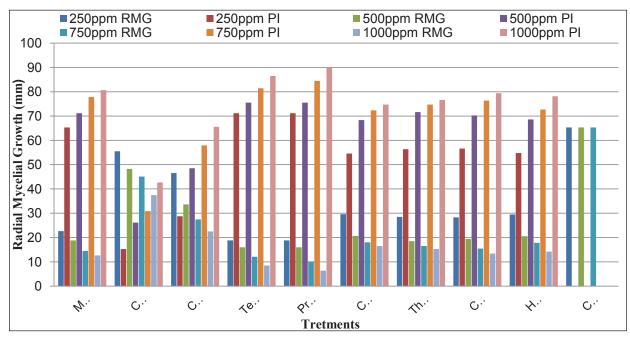


Fig. 1: Efficacy of different fungicides against Alternaria solani

et al. (2015) reported that Topsin M was more effective as compare to control., Ghazanfer *et al.* (2016), Kumar *et al.* (2017), Rani *et al.* (2017), Chapei *et al.* (2019), Sreenivasulu *et al.* (2019) evaluated that Tebuconazole was most effective followed by Propiconazole. Sudarshan G.K. (2020) and Sowmya and Chandra (2021). Who observed that Tebuconazole was the most effective fungicide followed by Difenconazole.

CONCLUSION

Among the fungicides Propiconazole was recorded

maximum mycelial growth inhibition of *A. solani,* other effective fungicides were Tebuconazole, Mancozeb, Captan + Hexaconazole, and Captan.

REFERENCES

- Anonymous. 2017. Horticulture Statistics at a Glance. NHB, Department of agriculture cooperation, Ministry of Agriculture, pp. 481.
- Bost, S. 2013. Foliar Diseases of Tomato. Plant Dis. SP277-W.
- Chaerani, R., Peralta, I.E., Knapp, S. and Spooner, D.M. 2005. New species of wild tomatoes (*Solanum* section *Lycopersicion*: Solanaceae) from Northern Peru. *Syst. Bot.*, **30**(2): 424-434.

IJAEB

- Chapei, S., Pongener, N. and Kangjam, V. 2019. Integrated Management of early blight of tomato (*Lycopersicum esculentum* Mill.) caused by *Alternaria solani* (Ellis and Martin) Jones and Grout. *Int. J. Curr. Microbiol. Appl. Sci.*, 8(5): 2428-2436.
- Chaurasia, A.K., Chaurasia, S., Chaurasia, S. and Chaurasia, S. 2013. Studies on the development of fruit rot of tomato caused by *Alternaria solani* (Ellis & Mart.) Jones & Grout. Biozone. *Int. J. Life Sci.*, **4**(6): 2713- 2716.
- Chohan, S., Perveen, R., Abid, M., Naz, M.S. and Akram, N. 2015. Morpho-physiological studies management and screening of tomato germplasm against *Alternaria solani* the causal agent of tomato early blight. *Int. J. Agric. Biol.*, **17**(1): 111-118.
- Ghazanfar Mu., Raza, W., Ahmed, K.S., Qamar, J., Haider, N. and Rasheed, M.H. 2016. Evaluation of different fungicides against *Alternaria solani* (Ellis & Martin) Sorauer cause of early blight of tomato under laboratory conditions. *Int. J. Zool.*, 5(1): 8-10.
- Gleason, M.L. and Edmunds, B.A. 2006. Tomato disease and disorders, http://www.extension.iastate.edu/store.
- Kumar, V., Singh, G. and Tyagi, A. 2017. Evaluation of different fungicides against Alternaria leaf blight of tomato (*Alternaria solani*). *Int. J. Curr. Microbiol. Appl. Sci.*, 6(5): 2343-2350.
- Maiero, M., Ng, T.J. and Barksdale, T.H. 1990. Genetic resistance to early blight in tomato breeding lines. *Horti. Sci.*, **25**(3): 344-346.
- Mathur, K. and Shekawat, K.S.1986. Chemical control of early blight in kharif sown tomato. *Indian J. Mycol. Pl. Pathol.*, **16**(2): 235-236.
- Mehta, R. 2017. History of Tomato (Poor Man's Apple). J. Humanit. Soc. Sci. (IOSR-JHSS), 22(8): 31-34.
- Momel, T.M. and Pemaezny, K.L. 2006. Florida plant disease management guide: Tomato. Florida Cooperation Extensive Service, institute of Food and Agriculture Sciences, http://edis.infas.ufl.edu.
- Nene, Y.L. and Thapliyal, P.N. 1993. Evaluation of fungicides. Fungicides in plant disease control. Oxford and IBH publishing company, New Delhi, pp. 531.
- Peralta, I.E., Knapp, S. and Spooner, D.M. 2005. New species of wild tomatoes (*Solanum* section *Lycopersicion*: Solanaceae) from Northern Peru. *Syst. Bot.*, **30**(2): 424-434.

- Rani, S., Singh, R. and Gupta, S. 2017. Development of integrated disease management module for early blight of tomato in Jammu. J. Pharmacogn. Phytochem., 6(2): 268-273.
- Sahu, D.K., Khare, C.P., Singh, H.K. and Thakur, M.P. 2013. Evaluation of newer fungicide for management of early blight of tomato in Chhattisgarh. *Int. J. Life Sci. Res.*, **8**(9): 1255-1259.
- Soni, R., Tanwar, V.K. and Yadav, S.M. 2017. Survey and Screening of Genotypes against *Alternaria solani* caused early blight of tomato in southern part of Rajasthan. *Chem. Sci. Rev. Lett.*, **6**: 1483-1489.
- Sowmya, V. and Chandra, R. 2020. *In vitro* and *in vivo* efficacy of chemical fungicides against early blight of tomato incited by *Alternaria solani* (Ell. & Mart.). *J. Pharmacogn. Phytochem.*, **10**(1): 833-837.
- Sreenivasulu, R., Reddy, M.S., Tomar, D.S., Subhash, M. and Reddy, B.B. 2019. Managing of Early Blight of Tomato Caused by *Alternaria solani* through Fungicides and Bioagents. *Int. J. Curr. Microbiol. Appl. Sci.*, 8(6): 1442-1452.
- Sudarshan, G.K., Nagaraj, M.S., Thammaiah N., Yogananada, S.B., Mallikarjuna Gowda, A.P. and Prasanna Kumar, M.K. 2020. *In vitro* Efficacy of Fungicides and Bioagents against Early Blight of Tomato caused by *Alternaria solani*. *Int. J. Curr. Microbiol. Appl. Sci.*, 9(9): 1490-1496.
- Tewari, R. and Vishunavat, K. 2012. Management of early blight (*Alternaria solani*) in tomato by integration of fungicide and cultural practices. *Int. J. Plant Prot.*, **5**: 201-206.
- Verma, K.P., Singh, S. and Gandhi, S.K. 2007. Variability among *Alternaria solani* isolates causing early blight of tomato. *Indian Phytopathol.*, 60(2): 180-186.
- Vincent, J.M. 1947. Distortion of fungal hyphae in the presence of certain inhibitors. *Nature*, **159**(4): 850-850.
- Voorrips R. 2006. Tomato early blight (*Alternaria solani*): The pathogen, genetics, and breeding for resistance. *J. Gen. Plant Pathol.*, **72**(6): 335–347.
- Walker, J.C. 1952. Disease of Vegetable Crops, 1st Ed. Mac Graw-Hill, New York, pp. 471-474.